

Michelle M. Hanna  
*To Be Assigned (Div. of Appl.*  
No. 09/984,664; Filed: October 30, 2001)

***Amendments***

***In the Drawings***

Please delete Figures 29A, 29B, and 29C.

Please substitute the informal drawings, Figures 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 30 and 31; with the attached formal drawings, Figures 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 and 30.

***In the Specification***

Please insert on page 1, after the title and inventor, and before the "BACKGROUND OF THE INVENTION" the following:

**CROSS REFERENCE TO RELATED APPLICATIONS**

The present application is a divisional of U.S. Application 09/984,664, filed October 30, 2001, and published on May 29, 2003 as patent application publication number US-2003-0099950; and incorporated by reference herein in its entirety.

Please delete paragraph [0069].

Please substitute paragraphs [0070] and [0071] for the following paragraphs [0070] and [0071]:

**[0070]** FIG. 29. Portion of the contig sequence of the CDKN2A gene. The sequence represents a small portion of the contig starting at 856630 nucleotides from the start of the contig sequence. The sequence represents a CpG island. Contig number: NT\_008410.4.

**[0071]** FIG. 30. Schematic representation of a “capture probe” to determine the methylation status of a specific gene. Oligonucleotide probes that are specific for a region near the CpG island of the target gene are immobilized onto a microtiter plate. The DNA of interest is added to the immobilized probe and bound to the capture probe. The DNA is then chemically modified to convert unmethylated C to T, and leave methyl-C unaffected. The converted DNA can then be amplified by an optional PCR step to further enhance the signal. A labeled CpG initiator is then added with an RNA polymerase and labeled nucleotide(s).

Please substitute paragraph [0244] with the following paragraph [0244]:

**[0244]** As the oligonucleotide product is generated, energy transfer occurs between TAMARA-SpApG and SF-UTP, which changes the wavelength at which TAMARA emits. If RNA polymerase or DNA is omitted from the reaction, there is no transfer of energy between the initiator and the terminator, and no change in the wavelength at which TAMARA emits.